

Cat Nr: MO-C032

Monolith NT™ Control Kit LabelFree

MicroScale Thermophoresis Grade

Control Kit for performing standard biomolecule interaction experiments using Monolith instruments including LabelFree detectors

CONTENT AND STORAGE

Monolith NT™ Control Kit LabelFree is shipped at room temperature.
Store at -20 °C upon arrival

Each kit contains material sufficient for 5 reactions

4 x Vial A (32 µg Protein)

4 x Vial B (25 µg Ligand)

4 x Vial C (500 µl of 1 x Buffer)

100 x 200 µl Tubes

Expiry date: see kit cover

INTRODUCTION

Control Kits contain biomolecules and a protocol for performing standard biomolecule interaction experiments using Monolith NT™ instruments. These kits are recommended

- when using Monolith instruments for the first time
 - for training new lab employees
 - for monitoring the correct performance of Monolith instruments
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SAMPLE PREPARATION PROTOCOL

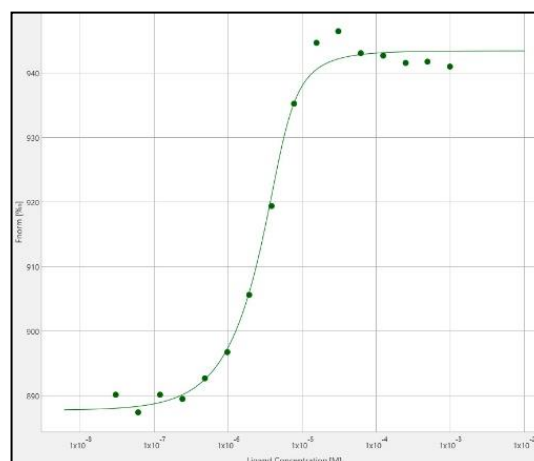
Please check the control software version available on your Monolith. For Monolith instruments equipped with MO.Control software, follow the instructions below. For Monolith instruments equipped with NTControl, please see next page for instructions.

PROTOCOL FOR USE WITH MO.CONTROL SOFTWARE

1. Add 25 µl of vial C (Buffer) to the vial B (Ligand) to dissolve the ligand. Let the solution sit for 5 minutes, then mix thoroughly by pipetting up and down. This will yield a 2 mM solution.
2. Add 200 µl of vial C (Buffer) to vial A (Protein) to dissolve the protein. Let the solution sit for 5 minutes, then mix thoroughly by pipetting up and down. DO NOT VORTEX. Make sure that the protein is dissolved completely. This will yield a 10 µM solution.
3. Start a Binding Affinity experiment. Fill in the Plan page as shown below. If supported by your Monolith instrument, it is also possible to use the auto-detect function for determining the optimal LED power setting. Then follow the instructions given by the software to run the experiment. On the Instructions page, the buffer is referred to as ligand buffer.

Target		Ligand	
Protein	<input type="text" value="Protein"/>	Ligand	<input type="text" value="Ligand"/>
Concentration of stock solution	<input type="text" value="10"/> <input type="text" value="µM"/>	Estimated K _d	<input type="text" value="optional"/> <input type="text" value="µM"/>
Concentration in this assay	<input type="text" value="5µM"/>	Concentration of stock solution	<input type="text" value="2000"/> <input type="text" value="µM"/>
Assay buffer	<input type="text" value="MST Buffer including 0.1% Pluronic"/>	Ligand in organic solvent like DMSO	<input type="checkbox"/>
Capillary	<input type="text" value="Monolith NT.Labelfree Zero Background Standard Treated Capillary (Z002)"/>	Ligand buffer in this assay	<input type="text" value="50.0%"/>
		Highest concentration in this assay	<input type="text" value="1000µM"/>
System settings			
Excitation Power	<input type="text" value="10 %"/>	MST-Power	<input type="text" value="Medium"/>
Auto-detect	<input checked="" type="checkbox"/>		

4. The interaction should show a K_d of around 1.5 µM.



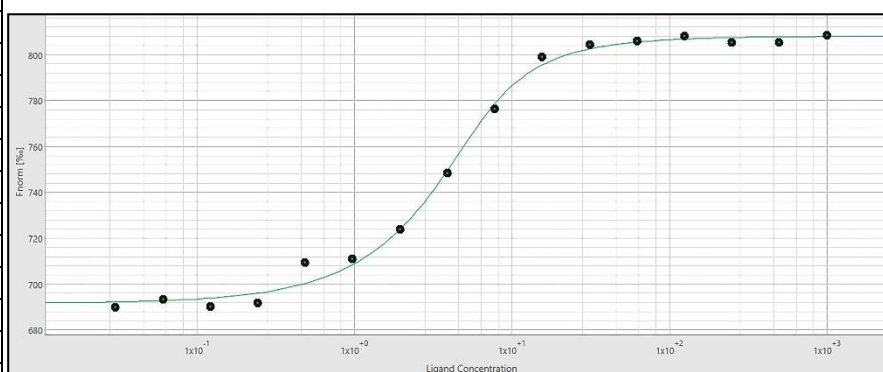
PROTOCOL FOR USE WITH NTCONTROL SOFTWARE

1. Add 25 μl of vial C (Buffer) to the vial B (Ligand) to dissolve the ligand. Let the solution sit for 5 minutes, then mix thoroughly by pipetting up and down. This will yield a 2 mM solution.
2. Prepare 16 reaction tubes. Add 20 μl from vial B (dissolved Ligand) to the first tube. Add 10 μl of vial C (Buffer) to the other 15 tubes. Transfer 10 μl with a clean tip from the first tube to the second tube and mix well by pipetting up and down. With a clean tip transfer 10 μl from the second tube to the third tube and mix well. Continue this 1-fold serial dilution until tube number 16. Remove and discard 10 μl from tube 16.
3. Add 200 μl of vial C (Buffer) to vial A (Protein) to dissolve the protein. Let the solution sit for 5 minutes, then mix thoroughly by pipetting up and down. DO NOT VORTEX. Make sure that the protein is dissolved completely. This will yield a 10 μM solution. Add 10 μl from vial A (dissolved Protein) to each tube containing Ligand dilution. Mix well by pipetting.
4. Load the instrument
Fill 16 Monolith™ NT.LabelFree Zero Background Standard Treated Capillaries with the prepared solutions and transfer the capillaries to the tray. Load the tray into the Monolith and start the measurement using the settings in the table below. First time users can find more detailed information on how to perform measurements in the instrument user manuals and starting guides.

LED power	10 %
MST power	40 %

5. Fit the data using the K_d model and fix the target concentration (TargetConc / Fluo. Conc.) to 5 μM . The interaction should show a K_d of around 1.5 μM .

Capillary	Concentration Ligand [μM]
1	1000.00
2	500.00
3	250.00
4	125.00
5	62.50
6	31.25
7	15.63
8	7.81
9	3.91
10	1.95
11	0.98
12	0.49
13	0.24
14	0.12
15	0.06
16	0.03



Contact

TECHNICAL SUPPORT

Please get in touch with us for specific questions concerning the product performance.

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PURCHASER NOTIFICATION

NanoTemper grants the buyer the non-transferable right to use the purchased product for research conducted by the buyer. The buyer cannot sell or otherwise transfer this product or its components for commercial purposes.

Limited warranty:

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